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(54) Title: RECOMBINANT RIBOSOMAL INHIBITOR PROTEIN (RIP) AND USE AS IMMUNOCONJUGATE			
(57) Abstract			
<p>The following description refers to a new RIP protein (SEQ ID No: 3) the cDNA sequence expressing same (SEQ ID No: 2), its preparation and use in the preparation of chemical and recombinant conjugates having anticancer properties.</p>			

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EE	Estonia						

RECOMBINANT RIBOSOMAL INHIBITOR PROTEIN (RIP) AND USE AS IMMUNOCONJUGATE

Field of the invention

5 The present invention relates to a new protein which - after being transferred into the cell by a suitable vector - is capable of inhibiting the activity of ribosomes. Therefore, it can be used as an anticancer and/or antiviral agent.

State of the art

10 As known, proteins extracted from filamentous fungi of the genus *Aspergillus*, such as α -sarcin, restrictocin and mitogillin, can inhibit protein synthesis by inactivating eukaryotic ribosomes. The nucleotide sequence of genomic DNA expressing said proteins is also known.

15 The antiviral activity of RIPs is related to the higher membrane permeability to the RIPs of virus-infected cells, with consequent injury to their ribosomes and consequent death of the infected cell. It follows that viral replication is interrupted. It has recently been disclosed that several RIPs inhibit HIV replication and that a RIP preparation, trichosantin (a protein extracted from the roots of *Trichosanthes kirilowii*) was used in phase I/II clinical studies [Byers, VS. et al., A phase I/II study of trichosantin treatment of HIV diseases, *AIDS*, 4, 1189-1196 (1990)].

20 With a view to obtain selectively cytotoxic molecules, several of the known RIPs were bound to proteic and non-proteic vectors capable of transferring them on specific target cell populations. Compounds with a specific cytotoxic action are most frequently prepared with monoclonal antibodies as protein vectors (immunotoxins). However, hormones, growth factors, lectins have also been used 25 as vectors for the treatment of cancer.

The protein that has been most widely used so far for the construction of immunotoxins is ricin chain A; however, several RIPs of type 1 (gelonin, PAP, saporin, momordin, bryodin, barley RIP) have been recently tested in the treatment, e.g., of tumours, autoimmune diseases, transplant rejections, 30 parasitoses, etc.

Since tumour cells are often toxin-resistant and all toxins used so far in therapeutic treatments induce an immune response in treated patients, the identification and purification to homogeneity of new RIPs are of great importance in all therapeutic applications and in particular in the generation of new immunotoxins.

5 **Summary of the invention**

The present invention relates to a new RIP protein SEQ ID No: 3, able to inhibit protein synthesis by inactivating ribosomes. Said RIP protein is herein referred to as clavin.

10 The present invention also concerns the nucleotide sequence SEQ ID No: 2, responsible of the expression of the new RIP protein clavin.

The present invention further includes the conjugates of the aforesaid protein with monoclonal antibodies, hormones, liposomes, growth factors, cytokines, transferrin and peptides, consisting of fragments of said proteins, obtainable by 15 chemical conjugation or by genic recombination techniques whenever applicable. It is another object of the present invention the Mgr6-clavin conjugate having amino acid composition SEQ ID No: 5, as well as the nucleotide sequence expressing it, corresponding to SEQ ID No: 4.

15 **Detailed description of the invention**

20 SEQ ID No: 1 reports the complete cDNA sequence (i.e., including non-coding 3' and 5' sequences) of clavin, in which:

1-279 = 5' UTR (5' fragment, untranslated);

280-360 = sequence encoding for hypothetical secretion sequence;

361-813 = sequence encoding for mature protein (clavin);

25 814-1011 = 3' UTR (3' fragment, untranslated) + polyA.

SEQ ID No: 2 refers to the cDNA sequence encoding for mature clavin.

SEQ ID No: 3 corresponds to clavin protein sequence.

SEQ ID No: 4 describes the nucleotide sequence encoding for the Mgr6-clavin immunotoxin produced in pRSET, in which:

30 1-108 = pRSET sequence containing the 6 histidines and the cleavage site for enterokinase;

109-861 = sequence encoding for ScFv (single Fv) of Mgr6
(109-462 = variable sequence of heavy chain, Bankit I.D. (Gene Bank) 54241,
access No. U 61494)
(463-507 = linker sequence)

5 508-861 = variable sequence of light chain, Bankit I.D. (Gene Bank) 54263,
access No. U 61495)
(862-1311 = sequence encoding for clavin).
SEQ ID IN No: 5 corresponds to the protein sequence of Mgr6-claim immunotoxin
produced in pRSET

10 The purified protein is >95% pure as shown by SDS/PAGE analysis, N-terminal
sequencing and reversed-phase HPLC. The protein molecular weight is approx
17kDa.

The following examples are conveyed for a better understanding of the protein
purification process according to the invention.

15 **cDNA isolation and sequencing**
Total RNA was extracted from *Aspergillus clavatus* IFO 8605 (Institute of
Fermentation, Osaka). mRNA was purified using the kits for total RNA and,
respectively, mRNA purification (Clontech). The two following primers were
synthesized:

20 3' α -primer: 5'-ACGTAAGCTTCTAATGAGAGCAGAGCTT-3' (SEQ ID No: 6)
5' α -primer: 5'-ACGTCTGCAGTGACCTGGACCTGCTTGAACG-3' (SEQ ID No: 7)
Primers were drawn on the basis of the known α -sarcin sequence by assuming a
high amino acid sequence homology with the toxin of *Aspergillus clavatus*.
The synthesis of cDNA with 5 μ g mRNA and 3' α -primer was carried out using an
25 appropriate synthesis kit (BRL). Part of the product obtained was added to a
reaction mixture for PCR containing Taq polymerase (USB) and the two aforesaid
primers; cDNA amplification for a total of 30 cycles was performed using a thermal
cyclizer for DNA (Perkin-Elmer).
Each cycle consisted of 1-min denaturation at 94°C, 1-min annealing at 42°C and
30 2-min extension at 72°C; in the final cycle extension at 72°C lasted 8 min.

Amplified cDNA corresponded to the cDNA of clavin, but contained the sequences imposed by 3'- α - and 5'- α -primers. Isolation of complete native cDNA of clavin was carried out by the RACE method.

On 3' end, RACE was performed according to Frohman, using the following 5 primer:

5'-GACTCGAGTCGACATCGA(T)₁₇-3' (SEQ ID No: 8), and the adjustment primer: 5'-GACTCGAGTCGACATCG-3' (SEQ ID No: 9).

Primer 5'-ACGTGGATCCTCTACAACCAGAAC-3' (SEQ ID No: 10), which refers to the codons for amino acids 23-29 of mature protein and bearing a restriction 10 site BamHI, was used as a gene-specific primer.

On 5' end, RACE was performed using the 5'-AmpliFINDER RACE Kit (Clontech), and primers

5'-TGAACCAGTGAGGATAG-3' (SEQ ID No: 11)

5'-ACGTCTGCAGGGCCTGTTCTCATA-3' (SEQ ID No: 12)

15 referring to the codons for amino acids 47-53 and 18-23 of mature protein were used as gene-specific primers; the latter primer also contains a restriction site PstI.

The various PCR products were purified, digested and subcloned in pUC19. Sequences were analysed using PC GENE software (Intelligenetics).

20 The complete cDNA sequence obtained is shown in Fig. 1. Said sequence contains an ORF encoding for a 177 amino acid polypeptide chain. The first 27 amino acids represent a signal peptide involved in secretion, while mature protein consists of the 150 amino acids shown in the figure.

Recombinant clavin heterologous expression

25 Vector pEZZ18 (Pharmacia) was used for recombinant clavin heterologous expression. Said vector directs the expression of fused proteins with a linking synthetic domain IgG (ZZ) based on staphylococcus protein A (Nilsson *et al.*, 1987). Clavin cDNA obtained by PCR with primers based on α -sarcin, as previously described, was re-amplified with 3' primer:

30 5'-GATCCTGCAGCGACCTGGACTTGCATGAACGAGCAGAAGAACCCAAAG-
ACC-3' (SEQ ID No: 13)

and with 3' primer: 5'-ACGTAAGCTTCTAATGAGAGCAGAGCTT-3' (SEQ ID No: 14)

to obtain the mature clavin native sequence, and cloned at restriction sites PstI-HindIII of vector pEZ18. To obtain pMRS116, fragment EcoRI-PstI was replaced

5 by linker B, which contains a sequence encoding for the cleavage site of factor Xa Ile-Glu-Gly-Arg, in addition to a residue Thr, inserted to preserve restriction site PstI. Linker B was obtained by annealing of the two oligonucleotides:

α-28: 5'-AATTCGATCGAAGGTCGTACTGCA-3' (SEQ ID No: 15)

α-29: 5'-GTACGACCTTCGATCG-3' (SEQ ID No: 16).

10 For clavin production, construction pMRS116 was propagated in *Escherichia coli* HB 101 [supE44, hsdS20(r_Bm_B)recA13, ara-14, proA2, lacY1, galK2, rpsL20, xyl-5, mtl-1] and cultured according to the producer's directions (Pharmacia).

Recombinant clavin purification

Culture supernatants were brought to pH 7.6 and added with 1 mM phenylmethylsulfonyl fluoride. Supernatants were injected into an IgG Sepharose Fast Flow column (Pharmacia) equalized with buffer (50 mM Tris/HCl, 150 mM NaCl and 0.05% Tween-20, pH 7.6). The column was first washed with said buffer and then with 5 mM ammonium acetate, pH 5.1; the fusion protein was eluted with 0.5 M ammonium acetate, pH 3.4, freeze-dried, dissolved to 2-8 mg/ml in 20 mM Tris/HCl, 100 mM NaCl, 1 mM CaCl₂, pH 8.0 and digested with Xa factor (Boehringer) at 23°C for 18-24 h in an enzyme/substrate ratio equal to 1:100.

20 Clavin was purified using a two-phase chromatographic process. In the first phase, the digestion mixture was injected into an S Sepharose Fast Flow column (Pharmacia) equalized with 20 mM sodium phosphate, pH 5.8, and eluted in NaCl gradient; the fusion protein left undigested and clavin were eluted in a single peak. The second phase was still performed on IgG Sepharose Fast Flow column: clavin was collected with the eluent, while the non-digested protein was eluted from the column with 0.5 M ammonium acetate, pH 3.4.

Immunotoxin synthesis and purification (chemical conjugate)

Clavin was derivatized with 40-molar excess ethyl S-acetyl-3-propionthioimidate at 4°C for 1 h to obtain an average molar ratio of ethyl S-acetyl-3-propionthioimidate groups to toxin molecule equal to 1.2.

5 Monoclonal antibody Mgr6, directed against the extracellular domain of ErbB2 and produced from hybridoma Mgr6-C4 MCB c # 762, deposited in the Interlab Cell Line Collection bank (CBA), an international deposit authority, was purified from ascitic liquid as described in Centis *et al.*, 1992. It was then derivatized with 10-molar excess 2-iminothiolane in ethanol at room temperature for 30 min, added
10 with 5,5'-dithio-bis(2-nitrobenzoic acid) to block the free -SH groups and obtain a groups/toxin molecule molar ratio equal to 1:6. The mixture was applied to a BioGel P-6DG column to remove all reagents.

15 Derivatized products were mixed using a 5-molar excess clavin, concentrated to a final volume of 1 ml, and added with 100 μ l of 0.5 M hydroxylamine, 12.5 mM EDTA, pH 7.2. The solution was stirred at 22°C for 14 h and at 4°C for additional 18 h. The reaction was interrupted by addition of 20 μ l of 200 mM N-ethylmaleimide.

20 The immunoconjugate was purified to homogeneity by ion exchange chromatography.

20 **Recombinant immunotoxin Mgr6-clavin**

Genic construction

25 The gene encoding for variable regions of monoclonal antibody Mgr6 was obtained by the "Recombinant Phage Antibody System" kit (Pharmacia) from mRNA of the antibody-producing hybridoma. Said procedure allows the obtainment of DNA encoding for ScFv (Single chain Fv), in which the sequences for the variable regions of heavy and light chains are joined by a linker sequence. The DNA for ScFv was then linked to clavin DNA to obtain the gene for immunotoxin, cloned in commercial vector pRSET (Introvigen). Figs. 4 and 5 show the nucleotide and, respectively, the amino acid sequence of the immunotoxin
30 inserted in pRSET.

The resulting plasmid has the following characteristics:

- a) the fusion protein gene is under the control of the T7 polymerase control;
- b) the resulting protein has at its N-terminal site an extension containing 6 histidines, usable for the purification with IMAC (immobilized metal affinity chromatography) and a cleavage site for enterokinase K.

5 **Recombinant immunotoxin expression in *E. coli***

Competent cells B834(DE3)pLys were transformed, plated on 10 LB agar plates containing ampicillin (100 µg/ml), and incubated at 37°C overnight.

Colonies were recovered in 500 ml LB culture medium containing ampicillin (100 µg/ml), glucose (0.5%) and MgSO₄ (1,62 mM), cultured at 37°C under stirring up

10 to OD₆₀₀ = 2.3/2.5, and, after addition of 1 l culture medium (LB, ampicillin, glucose, MgSO₄), amplified at 37°C under stirring up to OD₆₀₀ = 1.2.

Cells were centrifuged and resuspended in 1.5 l culture medium LB supplemented with ampicillin and induced by addition of IPTG (final 1 mM) under stirring at 37°C for 1.5 h. The cell pellet was recovered by centrifugation.

15 **Recombinant immunotoxin purification**

The pellet from 1.5 l culture medium was resuspended in 150 ml of 50 mM Tris-HCl, pH 8, and frozen. 30 ml aliquots were thawed out, sonicated (3 x 20 sec), and centrifuged at 160,00 rpm at 4°C for 30 min.

The resulting pellet was resuspended in 50 ml STET buffer (50 mM Tris-HCl, pH

20 8.5, 8% saccharose, 5% triton X-100, 50 mM EDTA) and the suspension was sonicated (3 x 45 sec) and centrifuged at 30,000 rpm for 20 min. The described washing procedure was repeated twice and twice again with 50 ml of 50 mM Tris-HCl, pH 8.5, and 100 mM NaCl.

Denaturation

25 The sample was resuspended in 30 ml buffer A (50 mM Tris-HCl, pH 8.0, 6 M Gu-HCl, 5 mM imidazole) and incubated at room temperature for at least 2 h.

Immunotoxin purification by IMAC

The sample was centrifuged at 120,00 rpm for 30 min. The supernatant was analysed by chelated metal affinity chromatography (IMAC) using 20 ml Ni⁺⁺-filled

30 chelating sepharose FF resin (Pharmacia).

The column was washed with 5 vol water, loaded with 5 vol of 0.1 M NiSO₄, washed with 5 vol water and equalized with 5 vol buffer A.

Once the sample had been injected, the column was washed with 5 vol buffer A.

Adsorbed proteins were eluted in step of pH using the following buffers:

5 50 mM Tris-acetate, pH 5.5; 6 M Gu-HCl (Buffer B);

50 mM Tris-acetate, pH 4.0; 6 M Gu-HCl (Buffer C).

The immunotoxin was eluted in buffer C.

Immunotoxin reduction

The sample obtained from IMAC (in a concentration of 1-2 mg/ml) was brought to

10 pH 8.3 with 1 M Tris base. 2 mM EDTA and final 300 mM DTT were added. The resulting product was incubated at room temperature for 3 h.

Immunotoxin refolding

The reduced sample was rapidly diluted (1:100) in the refolding buffer (50 mM Tris-HCl, pH 8.3, 0.5 M L-Arg, 2 mM EDTA, 4 mM GSSG, 2 mM DTT) and

15 incubated at 10°C for 60 h.

Immunotoxin concentration and dialysis

The sample (ca. 500 ml) was added with Tween-20 (final 0.005%) and concentrated by ultrafiltration through membrane Amicon YM 10.

Dialysis was carried out using a 10,000-cut-off membrane vs dialysis buffer (50

20 mM Tris-HCl, pH 8.0, 0.2 M NaCl, 0.005% Tween-20, 10% glycerol).

The resulting product was centrifuged at 120,00 rpm for 30 min and the supernatant containing the immunotoxin was recovered.

Inhibition of cell protein synthesis (chemical immunoconjugate)

The capacity of clavin and of immunotoxin Mgr6-clavin for inhibiting the protein

25 synthesis was measured on SKBr3 (ErbB2⁺ cells) and on MeWo (ErbB2⁺ cells) cultured in RPMI 1640 containing 10% FCS. The test was carried out essentially

as described by Casalini *et al.*, 1993.

The immunotoxin, the toxin or the monoclonal antibody were diluted in turn in the culture medium. Cells (1.2 x 10⁶) were incubated at 4°C for 3 h in polypropylene

30 test tubes, in 800 ul culture medium containing the appropriate concentrations of immunotoxin, toxin or monoclonal antibody alone. Control cells were incubated

only with the culture medium. Cells were then centrifuged, resuspended in a fresh culture medium and seeded in triplicate in 96-well plates (3×10^5 cells/well).

After incubation at 37°C for 48 h, the culture medium was removed and a fresh culture medium containing [3 H] proline (1uCi/well) was added. 48 h later, cells were washed and the amount of [3 H] proline incorporated was determined.

In various tests, clavin shows a dose/response effect with IC₅₀ values ranging from 0.1 to 1 μ M.

The cytotoxicity of the Mgr6-clavin conjugate is similar to that of ricin A bound to the same monoclonal antibody.

10 **Inhibition of cell protein synthesis (recombinant immunoconjugate)**

The capacity of recombinant immunotoxin Mgr6-clavin for inhibiting the protein synthesis was measured as already described for the chemical immunoconjugate. The IC₅₀ of recombinant immunotoxin ranges from 0.1 to 1 μ M, whereas antibody Mgr6 does not produce any effect. Therefore, clavin is a promising candidate for immunotoxin production.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

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- (H) TELEFAX: 02-54179920

(ii) TITLE OF INVENTION: A protein capable of inhibiting ribosomal activity, its preparation and use as a chemical or recombinant immunoconjugate, and the cDNA sequence expressing said protein.

(iii) NUMBER OF SEQUENCES: 16

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(vi) PRIOR APPLICATION DATA:

- (A) APPLICATION NUMBER: IT FI96A000155
- (B) FILING DATE: 27-JUN-1996

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1011 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Aspergillus clavatus
- (B) STRAIN: IFO8605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

AACCAGAAC AAAGGATATG TGGTGAGATT TGTGAGAAC CAGAACGCTT GAAAAAGAAA	60
ACAAAAGAGA GAAAAGTAAT CACCATCGAT GAGGATATTG TCTGACTCAG AGATCCAACG	120
AAATAATAGT CAACTTCGGA ATGCTTCAG TCGCCACAT CGAGCTGGT CATGGACTC	180
TCTCGAGTCG GCCAGAGCAC ATATAAAAGC TGCTAGATCC TCGCGGTTCT CCCAGGAAAA	240
CCCAAGATCG TGATCTCAAG CATCTTAACC ACATCCAAA TGGTGCAT CAAGAACCTC	300
GTCCCTGGTGC CCCTCACGGC CGTGACCGCC CTTCGCGATGC CTTCGCTCT CGAGGAGCGC	360
CGGGCGACCT GGACTTGCAT GAACGAGCAG AAGAACCCAA AGACCAACAA GTATGAGAAC	420
AAGGCCCTCC TCTACAAACCA GAACAATGCC GAGAGCAACG CCCACCCAGC GCCTCTCTCC	480
GACGGCAAGA CGGGTAGCAG CTATCCTCAC TGGTTCACCA ACGGCTACGA CGGGCATGGA	540
AAGATCTTCAGGGCGCAC GCCTTCAGA TGGGGAAATT CGGACTGCGA CGCCCTCTCCC	600
AAGCACACCA AGAATGGTGA TGGCAAGAAT GACCATTACC TGCTGGAGTT CCCAACATTC	660
CCCGATGGAC ACCAGTATAA TTTCGACTCG AAGAAGCCCA AGGAGGACCC CGGGCCGGCA	720
CGGGTCATCT ACACCTATCC TAACAAGGTG TTCTGCGGCA TTGTTGCCA CACGAGGGAG	780
AACCAGGGTG ACCTGAAGCT CTGCTCTCAT TAAATGGGCT TGCACAGGGTA TATAGTTGC	840
CATTGGTCGT TCTTCACCA CGGCTGATAC TATATGCAT TGGGAAGTGG GGGAGGGAGC	900
TGAATGTTTC ACATATGTTG GTGCAGAACT TGTTCATGT TATCTAGTCA ATCCCAAGTCT	960
CTCGCTTGTATCTATGCA TATTGCACTT CATTGCAAAA AAAAAAAAAA A	1011

(2) INFORMATION FOR SEQ ID NO: 2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 453 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: cDNA

- (iii) HYPOTHETICAL: NO

- (iv) ANTI-SENSE: NO

- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus
 - (B) STRAIN: IFO8605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GGGGCAGCTT	GGACCTTGCGAT	GAACGAGCG	AAGAACCCAA	AGACCAACAA	GTATGAGAAC	60
AAAGCGCTCC	TCTACAAACCA	GAACAAATGCC	GAGGACCAAGC	CCCACCCACGC	GCCTCTCTCC	120
GACGGCAAGA	CGGGTAGGAG	CTATCCTCAC	TGGTTACCCA	ACGGCTACGA	CGGGCGATGGA	180
AAAGATCTCA	AGGGCCGCAC	CCCCATCAG	TGGGAAATT	CGGACTGGGA	CGGGCCCTCCC	240
AAGCACAGCA	AGAATGGTGA	TGGCAAGAAT	GACCATTAAC	TGCTGGACTT	CCCAACATTG	300
CCCGATGGAC	ACCAGTATAA	TTTCGACTCG	AAGAAGGCCA	AGGAGGACCC	CGGGCCGGCA	360
CGGGTCACTCT	ACACCTATCC	TAACAAAGGTG	TTCTGCGGCA	TTGTTGCCCA	CACGAGGGAG	420
AAACAGGGTG	ACCTGAAGCT	CTGCTCTCAT	TAA			453

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 150 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Aspergillus clavatus*

(B) STRAIN: 100-665

(xii) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

Ala Ala Thr Trp Thr Cys Met Asn Glu Gln Lys Asn Pro Lys Thr Asn
1 5 10 15

Lys Tyr Glu Asn Lys Arg Leu Leu Tyr Asn Gln Asn Asn Ala Glu Ser
20 25 30

Asn Ala His His Ala Pro Leu Ser Asp Gly Lys Thr Gly Ser Ser Tyr
35 40 45

Pro His Trp Phe Thr Asn Gly Tyr Asp Gly Asp Gly Lys Ile Leu Lys
50 55 60

Gly Arg Thr Pro Ile Lys Trp Gly Asn Ser Asp Cys Asp Arg Pro Pro
65 70 75 80

Lys His Ser Lys Asn Gly Asp Gly Lys Asn Asp His Tyr Leu Leu Glu
 85 90 95

Phe Pro Thr Phe Pro Asp Gly His Gln Tyr Asn Phe Asp Ser Lys Lys
100 105 110

Pro Lys Glu Asp Pro Gly Pro Ala Arg Val Ile Tyr Thr Tyr Pro Asn
115 120 125

Lys Val Phe Cys Gly Ile Val Ala His Thr Arg Glu Asn Gln Gly Asp
130 135 140

Leu Lys Leu Cys Ser His
145 150

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1314 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Aspergillus clavatus
 - (B) STRAIN: IFO8605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

ATGCGGGGTT	CTCATCATCA	TCATCATCAT	GGTATGGCTA	GCATGACTGG	TGGACAGCAA	60
ATGGGTCTGGG	ATCTGTACGA	CGATGACGAT	AAGGATCGAT	GGGGATCCCA	GGTGCACTTG	120
CAGGAGTCTG	GGGCAGAGCT	TGTGAAGCCA	GGGGCCTCA	TCAAGTTGTC	CTGCACAGCT	180
TCTGGCTTCA	ACATTAAGA	CACCTATATG	CACTGGGTGA	AGCAGAGGCC	TGAACAGGGC	240
CTGGAGTGGG	TTGGAAGGAT	TGATCCTGCG	AATGGTAATA	CTAAATATGA	CCCGAAGTTC	300
CAGGGCAAGG	CCACTATAAC	AGCAGACACA	TCCTCCAAAC	CAGCCTACCT	GCAGCTCAGC	360
AGCCTGACAT	CTGAGGACAC	TGCCGTCTAT	TACTGTGCTA	GAGGAGATA	TGATTATCCT	420
TTTCCTTACT	GGGGCCAAGG	GACCTCGGTC	ACCGTCTCCT	CAGGTGGAGG	CGGTTCAAGGC	480
GGAGGGTGGCT	CTGGCGGTGG	CGGATCGTAC	ATCGAGCTCA	CTCAGTCTCC	AGCTTCCCTA	540
GCTGTATCTC	TGGGGCAGGG	GGCCACCATC	TCACTGCAGG	CCAGCCAAAG	TGTCAGTACA	600
TCTAGGTATA	GTTATATGCA	CTGGTACCAA	CAGAAACCAG	GACAGCCACC	CAARACTCCTC	660
ATCAAGTATG	CATCCAACCT	AGAATCTGGG	GTCCCTGCA	GGTTCAGTGG	CAGTGGGTCT	720
GGGGACAGACT	TCACCCCTCAA	CATCCATCT	GTGGGAGGAGG	AGGATACTGC	AACATATTAC	780
TGTCAGCACA	GTTGGGAGAT	TCCTCGGACG	TTCCGGTGGAG	GGACCAAGCT	GGAGCTGAAA	840
CGGGCGGGAT	CCCCGGAAAT	CGCAGCGACC	TGGACTTGCA	TGAACGAGCA	GAAGAACCCA	900
AAGACCAACA	AGTATGAGAA	CAAGCGCCTC	CTCTACAACC	AGAACAAATGC	CGAGAGCAAC	960
GCCCCACCAAG	CGCCCTCTCTC	CGACGCCAAG	ACCGGTAGCA	GCTATCCCTCA	CTGGTTCAACC	1020
AAACGGCTACG	ACGGCGATGG	AAAGATCCTC	AAGGGCCGCA	CGCCCATCAA	GTGGGGAAAT	1080
TCGGACTCGG	ACCGCCCTCC	CAAGCACAGC	AAGAATGGTG	ATGCGAAGAA	TGACCATTCAC	1140
CTGCTGGAGT	TCCCCAACATT	CCCCGATGGA	CACCACTATA	ATTCGACTC	GAAGAACGCC	1200
AAGGAGGACC	CCGGCCCGGC	ACGGGTCATC	TACACCTATC	CTAACAAAGGT	GTTCCTGGGC	1260
ATTTGTTGCC	ACACGAGGGG	GAACCAGGGT	GACCTGAAGC	TCTGCTCTCA	TTAG	1314

(2) INFORMATION FOR SEO ID NO: 5:

(1) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 437 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

(A) ORGANISM: *Aspergillus clavatus*
(B) STRAIN: IFO8605

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

Met Arg Gly Ser His His His His His Gly Met Ala Ser Met Thr
1 5 10 15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp
20 25 30

Arg Trp Gly Ser Gln Val Gln Leu Gln Glu Ser Gly Ala Glu Leu Val
35 40 45

Lys Pro Gly Ala Ser Val Lys Leu Ser Cys Thr Ala Ser Gly Phe Asn
50 55 60

Ile Lys Asp Thr Tyr Met His Trp Val Lys Gln Arg Pro Glu Gln Gly
65 70 75 80

Leu Glu Trp Ile Gly Arg Ile Asp Pro Ala Asn Gly Asn Thr Lys Tyr
85 90 95

Asp Pro Lys Phe Gln Gly Lys Ala Thr Ile Thr Ala Asp Thr Ser Ser
100 105 110

Asn Thr Ala Tyr Leu Gln Leu Ser Ser Leu Thr Ser Glu Asp Thr Ala
115 120 125

Val Tyr Tyr Cys Ala Arg Glu Tyr Asp Tyr Pro Phe Pro Tyr Trp
 130 135 140

Gly Gln Gly Thr Ser Val Thr Val Ser Ser Gly Gly Gly Gly Ser Gly
145 150 155 160

Gly Gly Gly Ser Gly Gly Gly Ser Tyr Ile Glu Leu Thr Gln Ser
165 170 175

Pro Ala Ser Leu Ala Val Ser Leu Gly Gln Arg Ala Thr Ile Ser Cys
180 185 190

Arg Ala Ser Gln Ser Val Ser Thr Ser Arg Tyr Ser Tyr Met His Trp
105 110 115

Tyr Gln Gln Lys Pro Gly Gln Pro Pro Lys Leu Leu Ile Lys Tyr Ala
310 315 320

Ser Asn Leu Glu Ser Gly Val Pro Ala Arg Phe Ser Gly Ser
255 256 257 258 259 260 261 262 263 264 265 266 267 268 269 270 271

Gly Thr Asp Phe Thr Leu Asn Ile His Pro Val Glu Glu Glu Asp Thr

Ala Thr Tyr Tyr Cys Gln His Ser Trp Glu Ile Pro Arg Thr Phe Gly
 260 265 270
 Gly Gly Thr Lys Leu Glu Leu Lys Arg Ala Gly Ser Pro Glu Phe Ala
 275 280 285
 Ala Thr Trp Thr Cys Met Asn Glu Gln Lys Asn Pro Lys Thr Asn Lys
 290 295 300
 Tyr Glu Asn Lys Arg Leu Leu Tyr Asn Gln Asn Asn Ala Glu Ser Asn
 305 310 315 320
 Ala His His Ala Pro Leu Ser Asp Gly Lys Thr Gly Ser Ser Tyr Pro
 325 330 335
 His Trp Phe Thr Asn Gly Tyr Asp Gly Asp Gly Lys Ile Leu Lys Gly
 340 345 350
 Arg Thr Pro Ile Lys Trp Gly Asn Ser Asp Cys Asp Arg Pro Pro Lys
 355 360 365
 His Ser Lys Asn Gly Asp Gly Lys Asn Asp His Tyr Leu Leu Glu Phe
 370 375 380
 Pro Thr Phe Pro Asp Gly His Gln Tyr Asn Phe Asp Ser Lys Lys Pro
 385 390 395 400
 Lys Glu Asp Pro Gly Pro Ala Arg Val Ile Tyr Thr Tyr Pro Asn Lys
 405 410 415
 Val Phe Cys Gly Ile Val Ala His Thr Arg Glu Asn Gln Gly Asp Leu
 420 425 430
 Lys Leu Cys Ser His
 435

(2) INFORMATION FOR SEQ ID NO: 6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

ACGTAAGCTT CTAATGAGAG CAGAGCTT

(2) INFORMATION FOR SEQ ID NO: 7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 31 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

ACGTCTGCAG TGACCTGGAC CTGCTTGAAC G

31

(2) INFORMATION FOR SEQ ID NO: 8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 35 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

GACTCGAGTC GACATCGATT TTTTTTTTTT TTTTT

35

(2) INFORMATION FOR SEQ ID NO: 9:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 17 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

GACTCGAGTC GACATCG

17

17

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 24 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

ACGTGGATCC TCTACACCCA GAAC

24

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

TGAACCAGTG AGGATAG

17

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

ACGTCTGCAG GCGCTTGTTC TCATA

25

(2) INFORMATION FOR SEQ ID NO: 13:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 51 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

GATCCTGCAG CGACCTGGAC TTGCATGAAC GAGCAGAAAG ACCCAAAGAC C

51

(2) INFORMATION FOR SEQ ID NO: 14:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 28 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

ACGTAAGCTT CTAATGAGAG CAGAGCTT

28

(2) INFORMATION FOR SEQ ID NO: 15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 24 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

AATTCGATCG AAGGTCGTAC TGCA

24

(2) INFORMATION FOR SEQ ID NO: 16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 16 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

GTACGACCTT CGATCG

16

Claims

1. Nucleotide sequence SEQ ID No: 2.
2. Protein capable of inactivating the ribosomal activity, the amino acid sequence SEQ ID No: 3.
3. Nucleotide sequence SEQ ID No: 4, encoding for the Mgr6-clavin immunotoxin.
4. Protein sequence SEQ ID No: 5 of Mgr6-clavin immunotoxin.
5. Conjugates, obtained by chemical conjugation or by genic recombination, of the protein as claimed in claim 2 with hormones, liposomes, monoclonal antibodies, growth factors, cytokines, transferrin and peptides consisting of fragments of said proteins.
6. The conjugates as claimed in claim 5, wherein the protein is conjugated with monoclonal antibodies.
7. The conjugates as claimed in claim 5, wherein the monoclonal antibody is Mgr6.
8. Pharmaceutical compositions containing, as active ingredient, the protein as claimed in claim 2 and/or the conjugates as claimed in any of claims 5 to 7, combined with suitable additives.
9. Use of the protein as claimed in claim 2 and/or of the conjugates as claimed in any of claims 5 to 7, or mixtures thereof, for the preparation of pharmaceutical preparations useful as anticancer and/or antiviral agents. A protein capable of inhibiting ribosomal activity, its preparation and use as a chemical or recombinant immunoconjugate, and the cDNA sequence expressing said protein.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP 97/03359A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C07K14/38 C07K19/00 C12N15/62 A61K47/48

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C07K C12N A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
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 Further documents are listed in the continuation of box C Patent family members are listed in annex

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- *A* document defining the general state of the art which is not considered to be of particular relevance
- *B* earlier document but published on or after the international filing date
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T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

Z document member of the same patent family

Date of the actual completion of the international search

24 November 1997

Date of mailing of the international search report

09-12- 1997

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Mateo Rosell, A.M.

INTERNATIONAL SEARCH REPORT

Intern. Appl. No.
PCT/EP 97/03359

C(Oneself) DOCUMENTS CONSIDERED TO BE RELEVANT		
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X	DATABASE WPI Week 9203 27 November 1991 Derwent Publications Ltd., London, GB; AN 92019323 XP002047697 & JP 03 266 986 A (OTSUKA SEIYAKU KOGYO KK), 27 November 1991 see abstract	1,2
Y	---	3,4
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A	EP 0 489 931 A (TORAY INDUSTRIES, INC.) 17 June 1992 see abstract and column 2, line 35-45.	1-9
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A	WO 95 11977 A (BRISTOL-MYERS SQUIBB COMPANY) 4 May 1995 see abstract and page 1, line 10-21 ---	1-9
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Intern. Appl. Application No.
PCT/EP 97/03359

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

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INTERNATIONAL SEARCH REPORT

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Intern. Application No

PCT/EP 97/03359

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